

# The Ammosamides: Structures of Cell Cycle Modulators from a Marine-Derived *Streptomyces* Species\*\*

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Marine-derived actinomycete bacteria are emerging as a valuable resource for bioactive natural products encompassing a variety of unique structural classes.<sup>[1]</sup> In our hands, early detection of cell growth inhibitors using in vitro cytotoxicity assays against the colon carcinoma cancer cell line HCT-116, followed by extensive mechanism of action studies, has proven to be an effective approach. As such, the HCT-116 assay has been instrumental in the identification of potentially important anticancer agents.<sup>[2]</sup>

In the course of our continued studies, *Streptomyces* strain CNR-698<sup>[3]</sup> was isolated from bottom sediments collected at a depth of 1618 meters in the Bahamas Islands in 2003. Cytotoxicity-guided (HCT-116) fractionation by C<sub>18</sub> flash chromatography and RP-HPLC of crude extract led to the isolation of ammosamides A (**1**) and B (**2**) as blue and red solids, respectively (3 and 4 mg L<sup>-1</sup>). Structure assignments for **1** and **2** proved to be particularly difficult due to their inherent insolubility (soluble only in dimethyl sulfoxide (DMSO)) and a lack of descriptive NMR signals, ultimately requiring the integration of NMR spectral analysis, mass spectrometry data, and single crystal X-ray diffraction studies.

High-resolution (ESI) mass spectrometric analysis of ammosamide A (**1**) indicated a molecular formula C<sub>12</sub>H<sub>10</sub><sup>35</sup>ClN<sub>5</sub>OS (*m/z* [M+H]<sup>+</sup>: 308.0303). The molecular weight of ammosamide B (**2**) was found to be 16 amu lower (*m/z* [M+H]<sup>+</sup>: 292.0604) consistent with the molecular formula C<sub>12</sub>H<sub>10</sub><sup>35</sup>ClN<sub>5</sub>O<sub>2</sub>. The UV/Vis spectrum of **1** was indicative of an unusually highly conjugated structure with absorptions at λ<sub>max</sub> = 580, 430, 350, and 290 nm.

Inspection of the <sup>1</sup>H NMR spectrum of **1** in [D<sub>6</sub>]DMSO revealed six singlets between δ = 6.0 and 9.0 ppm and one methyl singlet at δ = 4.03 ppm, while the <sup>13</sup>C NMR spectra revealed the presence of eleven sp<sup>2</sup> hybridized carbon atoms

and a single sp<sup>3</sup> hybridized carbon atom at δ<sub>c</sub> = 33.3 ppm (Table 1). The addition of D<sub>2</sub>O (20 μL) to the sample in [D<sub>6</sub>]DMSO resulted in the immediate disappearance of

Table 1: NMR spectral data for **1** and **2** ([D<sub>6</sub>]DMSO).

| Position              | Ammosamide A ( <b>1</b> )       |                               |                        | Ammosamide B ( <b>2</b> )       |                               |
|-----------------------|---------------------------------|-------------------------------|------------------------|---------------------------------|-------------------------------|
|                       | δ <sub>c</sub> <sup>[a,c]</sup> | δ <sub>H</sub> <sup>[d]</sup> | HMBC                   | δ <sub>c</sub> <sup>[b,c]</sup> | δ <sub>H</sub> <sup>[d]</sup> |
| 1a                    | 33.3                            | 4.03 (s)                      | C-2, C-8a              | 28.5                            | 3.59 (s)                      |
| 2                     | 177.2                           |                               |                        | 164.0                           |                               |
| 3                     | 116.5                           | 8.47 (s)                      | C-2, C-4<br>C-5b, C-4a | 115.3                           | 8.34 (s)                      |
| 4                     | 144.6                           |                               |                        | 144.7                           |                               |
| 4a                    | 166.0                           |                               |                        | 166.2                           |                               |
| 5b                    | 119.7                           |                               |                        | 119.0                           |                               |
| 7                     | 103.1                           |                               |                        | 104.5                           |                               |
| 8a                    | 110.5                           |                               |                        | 106.3                           |                               |
| CONH <sub>2</sub>     |                                 | 7.68 (brs)                    | C-4                    |                                 | 7.66 (brs)                    |
|                       |                                 | 8.92 (brs)                    | C-4a                   |                                 | 8.91 (brs)                    |
| NH <sub>2</sub> (C-6) |                                 | 7.16 (brs) <sup>[e]</sup>     | C-7                    |                                 | 6.73 (brs) <sup>[f]</sup>     |
|                       |                                 | 6.63 (s) <sup>[e]</sup>       |                        |                                 |                               |
| NH <sub>2</sub> (C-8) |                                 | 6.89 (s)                      | C-7, C-8a              |                                 | 6.18 (brs) <sup>[f]</sup>     |

[a] 75 MHz. [b] 125 MHz. [c] C-2a, C-5a, C-6, and C-8 (**1**: δ<sub>c</sub> = 132.6, 134.7, 136.8, 142.7 ppm; **2**: δ<sub>c</sub> = 130.6, 130.8, 132.4, 140.5 ppm) could not be unambiguously assigned. [d] 600 MHz. [e] At certain concentrations, these two one-proton signals coalesce to a two-proton signal at δ = 7.09 ppm. [f] Assigned by analogy to **1**.

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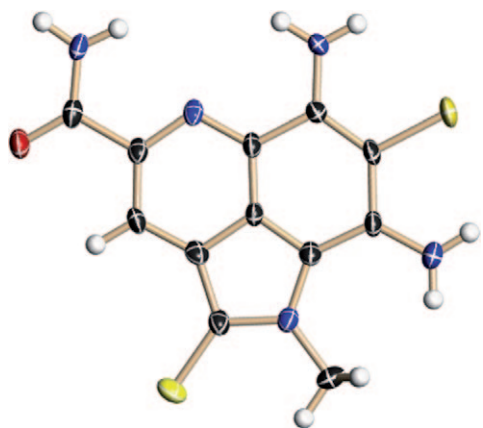
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<sup>1</sup>H NMR signals at δ = 7.16 (1H), 6.63 (1H), 6.89 ppm (2H) and the slower disappearance of singlets at δ = 8.92 (1H), and 7.68 ppm (1H) (less than 10 min). The exchangeable protons at δ = 7.16, 6.63 and 6.89 ppm were assigned as aromatic amines at C-6 and C-8 (based on HMBC correlations), while the slowly exchanging protons at δ = 8.92 and 7.68 ppm were assigned to a primary amide on the basis of COSY and HMBC correlations. The only non-exchangeable hydrogen atoms were the methyl singlet resonance at δ = 4.03 ppm and a one-proton singlet at δ = 8.47 ppm. The <sup>13</sup>C NMR spectrum of **1** indicated the presence of two carbonyl groups (δ<sub>c</sub> = 177.2 and 166.0 ppm), as well as two upfield sp<sup>2</sup> carbon atoms (δ<sub>c</sub> = 103.1 and 110.5 ppm). HMBC correlations between the downfield carbonyl (δ<sub>c</sub> = 177.2 ppm) and the proton methyl

singlet at  $\delta = 4.03$  ppm, we thought, defined an *N*-methyl amide, although a carbon chemical shift so far downfield would not be expected. In addition to correlations from the aromatic  $\delta = 8.47$  ppm singlet, the only other HMBC correlations were from the exchangeable protons at  $\delta = 7.16/6.63$  ppm to C-7 ( $\delta_c = 103.1$  ppm) and from  $\delta = 6.89$  ppm to C-7 and C-8a ( $\delta_c = 110.5$  ppm).

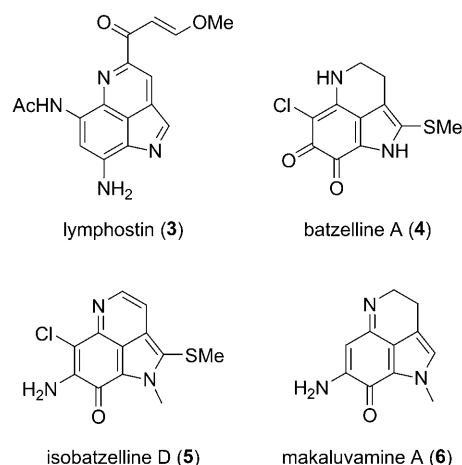
The spectral data for **1** suggested a highly unsaturated aza-aromatic metabolite possessing three rings. However, the lack of definitive NMR assignments that could be used to link these features forced us to concentrate efforts toward obtaining an X-ray crystal structure. We were fortunate to obtain small crystals of **1** by the slow diffusion of H<sub>2</sub>O into a saturated solution in DMSO.<sup>[4]</sup> The X-ray assignment of ammosamide A (**1**) is shown in Figure 1. Once X-ray data became clear, the spectral data for **1** could be assigned.



**Figure 1.** X-ray crystal structure of ammosamide A (**1**). Red O, blue N, yellow Cl, black C, white H.

The structure assignment of ammosamide B (**2**) followed from analysis of spectral data and chemical interconversion. Comparison of the C-2 carbonyl chemical shifts in **1** ( $\delta_c = 177.2$  ppm) and **2** ( $\delta_c = 164.0$  ppm) revealed a difference of 13 ppm, consistent with the typical <sup>13</sup>C chemical shift difference between a carbonyl and a thiocarbonyl (ca. 20 ppm).<sup>[5]</sup> In order to chemically confirm the presence of the thiolactam functionality, we used Lawesson's reagent [2,4-bis(*p*-methoxyphenyl)-1,3-dithiadiphosphetane-2,4-disulfide] to convert lactam **2** into thiolactam **1**.<sup>[6]</sup> The low yield of this reaction is likely attributable to the nucleophilic amines in **2**.<sup>[7]</sup> Exposed to air during storage, **1** was gradually converted to ammosamide B (**2**). Notably, the transformation could also be accomplished in 10 min, upon treatment of **1** with hydrogen peroxide in aqueous methanol.<sup>[8]</sup> This reactivity has been previously observed in other thioamide-containing compounds.<sup>[9]</sup>

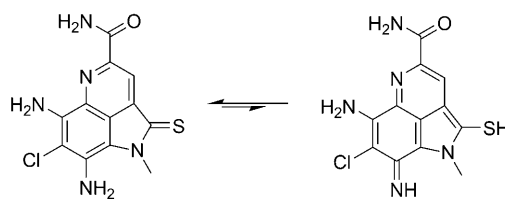
The structural similarities between the ammosamides and the microbial product lymphostin (**3**) are clear,<sup>[10]</sup> as is the relationship of the ammosamides to several sponge-derived pyrroloiminoquinone natural products, including batzelline A (**4**),<sup>[11a]</sup> isobatzelline D (**5**),<sup>[11b]</sup> and makaluvamine A (**6**)<sup>[11c]</sup> (Scheme 1). The sponge metabolites **4–6** possess different



**Scheme 1.** Related metabolites from bacteria and sponges.

patterns of Cl and NH<sub>2</sub> substitution and assume *p*-iminoquinone and *o*-quinone structures. The presence of an amino group at C-8 in the ammosamides results in a fundamentally different structure type in which the quinoline tautomer predominates. The pyrrole moiety in **3–6** is uniquely oxidized to the pyrrolidinone in ammosamide B (**2**). Finally, though methyl sulfides are present in **4** and **5**, ammosamide A (**1**) is the first natural product to contain a thio- $\gamma$ -lactam functionality.<sup>[12]</sup>

The fact that the ammosamides are highly colored (**1**:  $\lambda_{\max} = 580$  nm; **2**:  $\lambda_{\max} = 530$  nm), yet lack quinone or iminoquinone functionalities, leads to speculation about the electronic character and reactivity of these metabolites. The intense colors of these compounds could reflect a strong charge separation between the two six-membered aromatic rings due to the effects of electron-donating groups on the chlorine-containing ring and electron-withdrawing substituents on the pyridine ring. It is, conceptually, also explained by the potential for ammosamide A to exist in an equilibrium with its bis-iminoquinone tautomer (Scheme 2). Furthermore,



**Scheme 2.** Possible tautomeric form of ammosamide A (**1**).

in **1** and **2** the chlorine atom at C-7 is poised to engage in nucleophilic aromatic substitution with a suitable nucleophile, particularly when the molecule exists as its bis-iminoquinone tautomer.<sup>[13]</sup> This reactivity may be relevant to the molecule's interaction with its protein target.<sup>[14]</sup>

Ammosamides A (**1**) and B (**2**) exhibited significant in vitro cytotoxicity against HCT-116 colon carcinoma, each with IC<sub>50</sub> = 320 nM. These compounds also demonstrated

pronounced selectivity in a diversity of cancer cell lines with values ranging from 20 nM to 1  $\mu$ M, indicating a specific target mechanism of action. To explore the intracellular target of the ammosamides, ammosamide B (**2**) was converted to a highly fluorescent molecule by conjugation.<sup>[14]</sup> Treatment of HCT-116 colon carcinoma or HeLa cells with this fluorescent molecule produced immediate and irreversible labeling of a specific protein in the cellular cytosol. Using a cell and molecular biology approach, the target of the ammosamides was identified as a member of the myosin family, important cellular proteins that are involved in numerous cell processes, including cell cycle regulation, cytokinesis, and cell migration.

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